Vitamin D increases plasma renin activity independently of plasma Ca\(^{2+}\) via hypovolemia and \(\beta\)-adrenergic activity

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Atchison DK, Harding P, Beierwaltes WH. Vitamin D increases plasma renin activity independently of plasma Ca\(^{2+}\) via hypovolemia and \(\beta\)-adrenergic activity. Am J Physiol Renal Physiol 305: F1109–F1117, 2013. First published August 7, 2013; doi:10.1152/ajprenal.00010.2013.—1, 25-Dihydroxycholecalciferol (calcitriol) and 19-nor-1, 25-dihydroxyvitamin D\(_2\) (paricalcitol) are vitamin D receptor (VDR) agonists. Previous data suggest VDR agonists may actually increase renin-angiotensin activity, and this has always been assumed to be mediated by hypercalcemia. We hypothesized that calcitriol and paricalcitol would increase plasma renin activity (PRA) independently of plasma Ca\(^{2+}\) via hypercalcemia-mediated polycythemia, hypovolemia, and subsequent increased \(\beta\)-adrenergic sympathetetic activity. We found that both calcitriol and paricalcitol increased PRA threefold \((P < 0.01)\). Calcitriol caused hypercalcemia, but paricalcitol did not. Both calcitriol and paricalcitol caused hypercalcemia \((9- and 7-fold vs. control, \(P < 0.01)\) and polycythemia \((increasing 2.6- and 2.2-fold vs. control, \(P < 0.01)\). Paricalcitol increased renal calcium-sensing receptor \((CaSR)\) expression, suggesting a potential cause of paricalcitol-mediated hypercalcemia and polycythemia. Volume replacement completely normalized calcitriol-stimulated PRA and lowered plasma epinephrine by 43\% \((P < 0.05)\). \(\beta\)-Adrenergic blockade also normalized calcitriol-stimulated PRA. Cyclooxygenase-2 inhibition had no effect on calcitriol-stimulated PRA. Our data demonstrate that vitamin D increases PRA independently of plasma Ca\(^{2+}\) via hypercalcemia, polycythemia, hypovolemia, and increased \(\beta\)-adrenergic activity.

renin; PRA; calcium vitamin D; calcitriol; paricalcitol; calcium-sensing receptor \((CaSR)\); PTH; polycythemia; hypercalcemia; sympathetic activity; COX-2

1, 25-DIHYDROXYCHELALCALCIFEROL. (calcitriol) is a steroid hormone and is the active metabolite of the vitamin D system. Calcitriol exerts its effects via the vitamin D receptor \((VDR)\), a steroid nuclear receptor \((55)\). Calcitriol is known to play an important role in the regulation of calcium \((Ca^{2+})\) homeostasis. Calcitriol increases both renal and intestinal Ca\(^{2+}\) reabsorption \((27)\) and also permits parathyroid hormone \((PTH)\)-mediated bone resorption \((13)\), culminating in elevated plasma Ca\(^{2+}\). Calcitriol decreases plasma PTH levels indirectly via its effects on plasma Ca\(^{2+}\) \((8)\), as well as via direct effects on PTH transcription \((39)\).

Vitamin D has been previously shown to stimulate plasma renin activity \((PRA)\) \((32, 42)\). The mechanism by which this occurs remains unknown but has always been assumed to be due to the effects of hypercalcemia. Elevations in plasma Ca\(^{2+}\) are known to inhibit thick ascending limb \(Na^{+}-K^{+}-2Cl^{-}\) (NKCC) transport \((32)\) and induce polyuria and hypovolemia \((32)\), both of which could serve as powerful stimuli for increasing PRA. The polyuric effects of hypercalcemia are mediated by the calcium-sensing receptor \((CaSR)\) \((9)\), which impairs tubular transport in different nephron segments \((37, 50)\). Whether vitamin D could stimulate PRA independently of plasma Ca\(^{2+}\) is unknown. 19-nor-1,25-Dihydroxyvitamin D\(_2\) (paricalcitol) is a VDR agonist that suppresses plasma PTH, but unlike calcitriol, has minimal effects on plasma Ca\(^{2+}\) \((16, 43)\). To our knowledge, the effects of paricalcitol on renal Ca\(^{2+}\)-handling, polyuria, and concentrating defects have not been studied in depth. Paricalcitol could cause polycythemia and elevate PRA independently of plasma Ca\(^{2+}\) by upregulating the expression of the renal CaSR and causing hypercalcemia. Activating mutations of the CaSR are known to cause polycythemia, hypercalcemia, and hyperreninemia, in part by impairing collecting duct water transport. \((37, 46, 47, 52)\) mediated by aquaporin-2 \((AQP2)\). In support of this notion, paricalcitol is known to increase parathyroid gland CaSR mRNA \((54)\).

Vitamin D-induced hypercalcemia causes its polyuric effects in part through the increased production of prostaglandins \((38)\). Elevated Ca\(^{2+}\) can increase the expression and activity of cyclooxygenase-2 \((COX-2)\) in the renal medulla \((49)\), but whether COX-2 activity actually affects hypercalcemia-mediated polycythemia or PRA is unknown. It is well known that loop diuretics and dietary NaCl restriction increase PRA in part by increasing macula densa COX-2 activity and represent a major pathway for renin regulation \((18, 19)\). Whether hypercalcemia increases PRA via natriuresis-mediated elevations in macula densa COX-2 activity is unknown.

Additionally, hypercalcemia-induced polyuria could increase PRA due to reduced circulating volume, which is also a powerful stimulus for elevating PRA. Hypovolemia increases PRA in part due to an effect mediated by \(\beta\)-adrenergic activation \((11)\), and \(\beta\)-adrenergic receptors are integral for normal plasma renin levels \((20)\). In support of this, adrenal medullectomies and renal denervation partially impair the hypovolemia-mediated rise in PRA \((6)\). It is known that plasma catecholamine levels can be elevated in hypercalcemia \((41, 48)\). However, whether calcitriol-induced polyuria increases PRA via \(\beta\)-adrenergic stimulation is unknown.

The aim of this study was to determine whether vitamin D increases plasma renin independently of plasma calcium and to determine the mechanisms by which it does so. We hypothesized that treatment with calcitriol and paricalcitol will both increase PRA. We anticipated that paricalcitol would increase PRA independently of changes in plasma Ca\(^{2+}\) and will coincide elevated renal CaSR expression and hypercalcemia. Last, we anticipated that calcitriol would increase PRA via polycythemia-
induced hypovolemia mediated by elevated COX-2 activity and/or β-adrenoreceptor-mediated effects.

METHODS

Experimental Methods

Male Sprague-Dawley rats, weighing 200–250 g, singly housed in static caging, fed ad libitum, were used for all studies. Daily food consumption was measured by obtaining the remaining food in the cage each day. Water consumption was determined gravimetrically. Both were done to the nearest gram and milliliter, respectively. The day on which treatment protocols began was considered day 1. Rats were placed in metabolic caging on day 1 to equilibrate them before a 24-h urine collection performed on day 4. Rats were returned to their static caging on day 5, and in protocol I (see below) systolic blood pressure was measured with an automated tail-cuff system on which the rats had been trained previously three times (see below). In experiments in which PRA was quantified, rats were euthanized on day 6 by decapitation to collect PRA samples unaffected by anesthesia. The first 3 s of free-flowing trunk blood were collected for PRA analyses in chilled tubes containing 50 μl of 3.8% EDTA in 0.9% NaCl. The collection of only the first 3 s of blood is essential to ensure that PRA analyses are not contaminated by baroreceptor-elevated PRA levels from the decapitation. Additional blood samples were collected from free-flowing trunk blood using either 50 μl of 3.8% EDTA (Sigma-Aldrich, St. Louis, MO) or sodium heparin (Sagent Pharmaceuticals, Schaumberg, IL) as anticoagulants. Blood samples were spun at 1,164 g at 4°C for 15 min, and the plasma was aspirated and stored at −20°C until further analysis. The left kidney of each rat was rapidly excised and weighed.

In experiments in which renal CaSR mRNA was quantified, rats were injected with 50 mg/kg Nembutal (pentobarbital sodium ip, Ovation Pharmaceuticals, Deerfield, IL) on day 6. The peritoneal cavity was opened with a midline incision. Under sterile conditions, the left kidney was quickly exposed, clamped, excised, and the capsule was removed. The kidney was bisected and immediately placed in Tri-Reagent (Molecular Research Center, Cincinnati, OH). After homogenization of the kidneys, they were centrifuged at 16,000 g for 10 min at 4°C. The suprenatant was aspirated and stored at −80°C until RNA was isolated. Rats were euthanized via aortic transection and bilateral pneumothorax.

Animal Welfare Assurance

All procedures were approved by the Henry Ford Health System Institutional Animal Care and Use Committee and adhered to the guiding principles in the care and use of experimental animals in accordance with the National Institute of Health (NIH) guidelines. Henry Ford Hospital operates an AALAC-certified animal care facility.

Analyses

**PRA.** PRA was analyzed by generation of ANG I (ng ANG I·ml·h⁻¹·min⁻¹) using a Gamma Coat RIA kit (DiaSorin, Stillwater, MN) as previously described and according to the manufacturer’s instructions (3–5).

**Plasma ionized Ca²⁺, plasma Na⁺, urinary Ca²⁺, and Na⁺ quantification.** Plasma Ca²⁺ and Na⁺ were measured using a NOVA-8 electrolyte analyzer (NOVA Biomedical, Waltham, MA). Urinary Ca²⁺ and PO₄³⁻ were measured with colorimetric (Biovision, Mountain View, CA) assay kits according to the manufacturer’s instructions using a colorimetric plate reader (Tittertek, Huntsville, AL). Absorbance was measured at 570 and 620 nm, respectively, and values were analyzed with Multiskan Ascent. Urinary Na⁺ was measured with a NOVA-I electrolyte analyzer (NOVA Biomedical). The concentrations of Ca²⁺, PO₄³⁻ and Na⁺ in the urine were multiplied by the 24-h urinary volume to achieve the amount of Ca²⁺ or Na⁺ excreted in 24 h.

**Plasma PTH quantification.** In protocols 1, 2, and 3, plasma PTH 1–84 was quantified using an enzyme-linked immunnoassay (Alpcos Diagnostics, Salem, NH) according to the manufacturer’s instructions as described previously (3–5).

**Plasma and urinary creatinine quantification and creatinine clearance calculation.** Plasma and urinary creatinine were determined using a colorimetric assay (BioAssay Systems, Hayward, CA). Creatinine clearance was calculated by multiplying the concentration of urinary creatinine by the 24-h urinary volume, dividing by the plasma creatinine concentration, and then correcting the units of time for clearance to milliliters per minute. Last, clearance values were normalized per gram of kidney weight. The units for creatinine clearance are milliliters per minute per gram kidney weight.

**Urine and plasma osmolality.** Urine and plasma osmolality were measured using a model 3300 Advanced Micro Osmometer (Advanced Instruments, Norwood, MA).

**Tail-cuff plethysmography.** In protocol I, systolic blood pressure was measured noninvasively using a computerized tail-cuff system (model 1231, ITC, Woodland Hills, CA). Rats were trained over 3 days before systolic blood pressure measurement. Three systolic blood pressure measurements were taken from each rat, and a mean value was calculated for statistical analyses.

**Plasma epinephrine.** Plasma epinephrine was determined using a commercially available ELISA kit (Rocky Mountain Diagnostics, Colorado Springs, CO) according to the manufacturer’s instructions.

**Real-time quantitative RT-PCR.** Quantification of CaSR mRNA was performed by quantitative real-time RT-PCR using a SYBR green method. Custom rat-specific primers from TIB Molbiol (Adelphia, NJ) were used for all of the PCRs. The primer sequences for CaSR, NCC2, and AQP2 are as follows, respectively: forward 5’-ctgaga-ggaagaacagca-3’, reverse 5’-ttccatgcgtgtgcact-3’; forward 5’- ggctcatagcc-3’, reverse 5’-ggtagttgcttgcctc-3’; and forward 5’-gcactctggatc-3’, reverse 5’-ccgtatcacactgc-3’. Real-time RT-PCR was performed as follows: 1 μg of DNase-treated total RNA sample was reverse transcribed using random primers and Omniscript reverse transcriptase (Qiagen, Valencia, CA) in a total volume of 20 μl for 1 h at 37°C followed by an inactivation step of 95°C for 5 min. Two microliters of the reverse transcription reaction was then amplified in a Roche version 2.0 LightCycler PCR instrument (Roche, Indianapolis, IN) using SYBR green dye (SA Biosciences, Frederick, MD) and specific primers. Reactions were set up in a final volume of 20 μl, which contained 2 μl of sample, 1 μmol/l of each of the primers, and 10 μl of 2× SYBR green PCR mix. After an initial “hot start” at 95°C for 10 min, amplification occurred by denaturation at 95°C for 15 s, annealing at 58°C for 35 s, and extension at 72°C for 35 s, for a total of 30–40 cycles. At the end of PCR cycling, melting curve analyses were performed. A relative quantitation method [ΔΔCt] (53) was used to evaluate expression of CaSR. RT-PCR of GAPDH was used for normalization of all data.

Experimental Protocols

**Protocol 1: effect of calcitriol and paricalcitol on PRA and CaSR, NCC2, and AQP2.** We administered 100 ng of calcitriol once per day (ip) delivered in 100 μl of DMSO. Vehicle control-treated rats were treated with matching volumes of DMSO. This dose of calcitriol was chosen due to the ability of a similar dose to decrease renal renin mRNA in mice (16, 34). To test whether calcitriol was increasing PRA due to its stimulatory effects on plasma Ca²⁺, we administered a “noncalcemic” analog of vitamin D, paricalcitol, which binds to and activates the VDR (43). Paricalcitol capsules (Zemplar, Abbott Pharmaceuticals) were purchased at the Henry Ford Hospital Pharmacy and were not gifts. Paricalcitol was drained from capsules using a 26-gauge needle and suspended in DMSO. Paricalcitol was injected 100 ng/day (ip) once daily. This dose of paricalcitol was chosen due
to its inability to increase plasma Ca\(^{2+}\) (15, 16, 43, 56) and also its reported ability to decrease PRA in spontaneously hypertensive rats (22) and renal renin mRNA in mice (26). The numbers were \(n = 8\) in the control and calcitriol groups and \(n = 7\) for the paricalcitol group.

Additionally, to test whether the polyuria induced by paricalcitol was due to changes in CaSR, NKCC2, or AQ2P expression, rats were treated with 100 ng/day calcitriol (\(n = 5\)), paricalcitol (\(n = 5\)), or DMSO (control, \(n = 5\)). On day 6, rats were euthanized as described in Experimental Methods. Quantitative RT-PCR was performed on RNA from whole-kidney homogenates to test for changes in CaSR, NKCC2, or AQ2P expression.

Protocol 2: effects of volume replacement on calcitriol-stimulated PRA. To test whether calcitriol-induced hypercalcemia increased PRA due to its effects on polyuria and hypovolemia, we performed volume replacement in calcitriol-treated rats. Rats were treated with 100 ng/day of calcitriol (ip). One group of calcitriol-treated rats received 25 ml of sterile 0.9% NaCl daily, in divided doses, subcutaneously, except on the final day, on which the rats received all 25 ml 5 h before euthanasia. This dose of saline was chosen to make the rats euolemic because the calcitriol-treated rats in protocol 1 weighed \(\sim 25\) g less than the vehicle control-treated rats at the end of the study. Numbers were \(n = 8\) for the control group and \(n = 7\) for the calcitriol+0.9% NaCl group. Additionally, to demonstrate that calcitriol clearly increased plasma epinephrine levels compared with control, we sampled plasma from \(n = 5\) control rats to include in Fig. 4.

Protocol 3: effect of \(\beta\)-adrenoreceptor inhibition on calcitriol-stimulated PRA. To test whether calcitriol increased PRA due to elevated \(\beta\)-adrenergic activity, we administered the propranolol (Sigma-Aldrich) to calcitriol-treated rats. Rats received 100 ng of calcitriol/day. Starting on day 1, one group of calcitriol-treated rats received 4 mg/kg propranolol daily in divided doses (ip) delivered in 100 \(\mu\)l of sterile 0.9% NaCl (pH = 2). This dose was based on a similar dose shown to inhibit PRA in hypovolemic states (11). A second group received calcitriol and the vehicle for propranolol. A third group received propranolol in the absence of calcitriol, and a final group received both the vehicles for propranolol and calcitriol. Numbers for all groups were \(n = 10\).

Protocol 4: effect of COX-2 inhibition on calcitriol-stimulated PRA. To test whether calcitriol-induced hypercalcemia increased PRA due to increased COX-2 activity, we administered the COX-2-selective inhibitor NS-398 (Cayman Chemical, Ann Arbor, MI) to calcitriol-treated rats. Rats received 100 ng of calcitriol/day. Starting on day 3, one group (\(n = 18\)) received 10 mg/kg NS-398/day, in divided doses, in 50 \(\mu\)l of DMSO (ip). This dose of NS-398 was chosen due to its previously demonstrated ability to inhibit COX-2 (26). The calcitriol control group (\(n = 18\)) received calcitriol and the vehicle for NS-398 (DMSO). Additionally, extra groups of rats receiving NS-398 in the absence of calcitriol (\(n = 8\)), or only DMSO (\(n = 8\)) were included.

**Statistical Analyses**

Single intergroup comparisons between two groups were performed with a Student’s t-test (protocols 1 and 2). When multiple comparisons with one differing factor were performed, one-way ANOVA with a Student-Newman-Keuls post hoc test was performed (protocols 1 and 2). Two-way ANOVA was performed on nested designs where we tested for two-way interactions (protocols 3 and 4). Individual comparisons using a one-way ANOVA design with a Student-Newman-Keuls test was used to examine individual pairwise comparisons post hoc. Pairwise comparisons tested for differences between each treatment within the two factors separately. The treatments within one factor were compared among themselves without regard to the second factor. For simplicity of presentation, all data are presented as means ± SE. In all cases, \(P < 0.05\) was considered statistically significant. The absolute values for PRA, epinephrine, and CaSR/GAPDH, NKCC-2/GAPDH, and AQ2P/GAPDH mRNA ratios are found in the text. All other absolute values are found in tables.

**RESULTS**

**Protocol 1: effect of calcitriol and paricalcitol on PRA and the CaSR, NKCC2, and AQ2P.** To test whether calcitriol could increase PRA, we administered 100 ng/day of calcitriol. Additionally, to test whether the increases in PRA were independent of changes in plasma Ca\(^{2+}\), we administered 100 ng/day of paricalcitol (a noncalcemic VDR agonist) to an additional group. Calcitriol increased PRA from 3.1 ± 0.3 to 10.3 ± 1.0 ng ANG 1-ml\(^{-1}\)-h\(^{-1}\) \((P < 0.001,\) Fig. 1). Paricalcitol also similarly increased PRA to 8.8 ± 2.0 ng ANG 1-ml\(^{-1}\)-h\(^{-1}\) \((P < 0.01,\) Fig. 1).

Ionized Ca\(^{2+}\) was significantly higher in calcitriol-treated rats by 23% (Table 1), but plasma Ca\(^{2+}\) in the paricalcitol group did not differ from controls (Table 1), demonstrating that vitamin D can increase PRA in the absence of hypercalcemia. Confirming the successful administration of calcitriol and paricalcitol in these rats, plasma PTH was greatly suppressed in both the calcitriol- and paricalcitol-treated groups compared with control (Table 1). The 24-h urinary volume more than doubled in both the calcitriol- and paricalcitol-treated groups (Table 1). Urinary Na\(^{+}\) excretion was higher in the paricalcitol group compared with both control and calcitriol (Table 1). Urinary Ca\(^{2+}\) excretion was significantly higher, and urine osmolality significantly lower in both the calcitriol- and paricalcitol-treated groups compared with controls (Table 1) without significant changes in the creatinine clearance. Urinary PO\(_4\)\(^{3-}\) excretion increased in both the calcitriol and paricalcitol groups, but was significantly higher in the calcitriol group compared with paricalcitol (Table 1). The final body weight (at the end of treatment) was lower in the calcitriol-treated group, but not the paricalcitol group (Table 1). Food consumption was slightly, but significantly lower in the calcitriol-treated group, but not the paricalcitol-treated group (Table 1). Thus calcitriol and paricalcitol both increased PRA, but this response was dissociated from changes in plasma Ca\(^{2+}\). However, rats in both groups were characterized by a significant polyuria, hypercalciumia, and an apparent renal concentrating defect.

To test a potential mechanism for calcitriol- and paricalcitol-induced hypercalciumia and polyuria, we examined the effects of paricalcitol on whole-kidney homogenate CaSR, NKCC2, and AQ2P expression. Calcitriol decreased renal CaSR/GAPDH mRNA expression from 1.21 ± 0.17 to 0.48 ± 0.17...
Vehicle control-treated rats at the end of the study. Compared to euvolemia, this dose of saline was chosen because the polyuric, polyvinyl chloride to calcitriol-treated rats daily. To obtain due to polyuria-mediated hypovolemia, we administered 25 ml stimulated PRA.

Twenty-four-hour urine volume and urinary Na

Values are means ± SE. *P < 0.05 vs. control. **P < 0.01 vs. control. ***P < 0.001 vs. control. †P < 0.05 vs. calcitriol. ††P < 0.01 vs. calcitriol. †††P < 0.001 vs. calcitriol.

(2A, P < 0.05). Paricalcitol increased renal CaSR/GAPDH mRNA expression to 2.06 ± 0.29 (Fig. 2A), significantly greater than control (P < 0.05) and calcitriol (P < 0.001). Calcitriol also significantly decreased renal NKCC2/GAPDH and mRNA from a control level of 1.07 ± 0.18 to 0.40 ± 0.09 (Fig. 2B, P < 0.05). NKCC2/GAPDH mRNA expression was 1.30 ± 0.25 in paricalcitol-treated rats. This was significantly higher than calcitriol (Fig. 2B, P < 0.05), but did not differ from control. Neither calcitriol nor paricalcitol significantly affected AQP2/GAPDH mRNA expression (Fig. 2C).

Protocol 2: effects of volume replacement on calcitriol-stimulated PRA. To test whether calcitriol was increasing PRA due to polyuria-mediated hypovolemia, we administered 25 ml of sterile 0.9% NaCl to calcitriol-treated rats daily. To obtain euvoolemia, this dose of saline was chosen because the polyuric, calcitriol-treated rats in protocol 1 weighed ~25 g less than the vehicle control-treated rats at the end of the study. Compared with calcitriol-treated rats (9.2 ± 1.8 ng ANG I·ml

Table 1. Effects of calcitriol and paricalcitol on plasma and urinary parameters, body weight, food and water intake, and systolic blood pressure

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>Calcitriol</th>
<th>Paricalcitol</th>
</tr>
</thead>
<tbody>
<tr>
<td>Plasma Na⁺, mmol/l</td>
<td>140 ± 0.2</td>
<td>139 ± 0.2</td>
<td>139 ± 0.6</td>
</tr>
<tr>
<td>Plasma ionized Ca²⁺, mmol/l</td>
<td>1.24 ± 0.01</td>
<td>1.52 ± 0.03***</td>
<td>1.23 ± 0.02†††</td>
</tr>
<tr>
<td>PTH, pg/ml</td>
<td>43.5 ± 2.3</td>
<td>0.7 ± 0.3***</td>
<td>6.4 ± 3.2***</td>
</tr>
<tr>
<td>Plasma osmolality, mosmol/kgH₂O</td>
<td>304 ± 4</td>
<td>305 ± 4</td>
<td>300 ± 3</td>
</tr>
<tr>
<td>24-h Urine volume, ml/24 h</td>
<td>11.3 ± 1.0</td>
<td>29.3 ± 3.4***</td>
<td>24.6 ± 3.2**</td>
</tr>
<tr>
<td>Urinary Na⁺ excretion, mmol/24 h</td>
<td>1.48 ± 0.10</td>
<td>1.73 ± 0.06</td>
<td>2.29 ± 0.23**†</td>
</tr>
<tr>
<td>Urinary Ca²⁺ excretion, mg/24 h</td>
<td>1.31 ± 0.19</td>
<td>11.7 ± 2.5**</td>
<td>9.2 ± 2.6*</td>
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<tr>
<td>Urinary PO₄³⁻ excretion, mmol/24 h</td>
<td>0.72 ± 0.06</td>
<td>1.78 ± 0.16***</td>
<td>1.25 ± 0.10**††</td>
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<tr>
<td>Creatinine clearance, ml·min⁻¹·g kidney wt⁻¹</td>
<td>0.81 ± 0.07</td>
<td>0.89 ± 0.07</td>
<td>0.66 ± 0.05</td>
</tr>
<tr>
<td>Urine osmolality, mosmol/kgH₂O</td>
<td>1.623 ± 108</td>
<td>786 ± 92***</td>
<td>1.045 ± 137**</td>
</tr>
<tr>
<td>Basal body weight, g</td>
<td>236 ± 5</td>
<td>234 ± 5</td>
<td>244 ± 4</td>
</tr>
<tr>
<td>Final body weight, g</td>
<td>274 ± 2</td>
<td>249 ± 5**</td>
<td>272 ± 4†††</td>
</tr>
<tr>
<td>Food consumed, g/day</td>
<td>17.9 ± 1.0</td>
<td>14.4 ± 0.8*</td>
<td>17.1 ± 0.5†</td>
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<tr>
<td>H₂O consumed, ml/day</td>
<td>45 ± 3</td>
<td>53 ± 5</td>
<td>51 ± 2</td>
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<tr>
<td>Systolic blood pressure, mmHg</td>
<td>128 ± 9</td>
<td>132 ± 8</td>
<td>130 ± 4</td>
</tr>
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Fig. 2. A: effects of calcitriol and paricalcitol on whole-kidney calcium-sensing receptor (CaSR) mRNA. Calcitriol decreased CaSR mRNA. Paricalcitol increased CaSR mRNA compared with control and calcitriol. Values are means ± SE and are corrected by GAPDH expression. *P < 0.05 vs. control. ††P < 0.001 vs. calcitriol. B: effects of calcitriol and paricalcitol on whole-kidney Na-K-2 Cl cotransporter (NKCC2) mRNA. Calcitriol decreased NKCC2 mRNA vs. both control and paricalcitol. Values are means ± SE and are corrected by GAPDH expression. *P < 0.05 vs. control. †††P < 0.001 vs. calcitriol. C: effects of calcitriol and paricalcitol on whole-kidney aquaporin-2 (AQP2) mRNA. Neither paricalcitol nor calcitriol affected AQP2 mRNA expression. Values are means ± SE and are corrected by GAPDH expression.
calcitriol was significantly lower in calcitriol vs. their respective controls (Table 3). Interestingly, body weight
Urinary Na
24-h Urine volume, ml/24 h 35.6
/H11006
Plasma osmolality,
Urine osmolality, mosmol/kgH2O 677
/H11006
Plasma epinephrine, pg/ml 1,695.0
/H11006
H2O consumed, ml/day 69
/F1113
Food consumed, g/day 19.4
/H11006
Final body weight, g 270
/H11006
Basal body weight, g 258
/H11006
Plasma Ca2+
Calcitriol Calcitriol

Parameter
Calcitriol Calcitriol+ 0.9%NaCl

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Calcitriol</th>
<th>Calcitriol+ 0.9% NaCl</th>
</tr>
</thead>
<tbody>
<tr>
<td>Plasma Na+ (mmol/l)</td>
<td>139 ± 0.4</td>
<td>140 ± 0.7</td>
</tr>
<tr>
<td>Plasma Ionized Ca2+ (mmol/l)</td>
<td>1.39 ± 0.03</td>
<td>1.37 ± 0.05</td>
</tr>
<tr>
<td>PTHm pg/ml</td>
<td>35.2 ± 12.6</td>
<td>5.6 ± 3.5</td>
</tr>
<tr>
<td>Plasma osmolality, mosmol/kgH2O</td>
<td>304 ± 1</td>
<td>302 ± 2</td>
</tr>
<tr>
<td>24-h Urine volume, ml/24 h</td>
<td>35.6 ± 3.8</td>
<td>55.9 ± 4.1***</td>
</tr>
<tr>
<td>Urinary Na+ excretion, mmol/24 h</td>
<td>1.70 ± 0.10</td>
<td>6.23 ± 0.63***</td>
</tr>
<tr>
<td>Urinary Ca2+ excretion, mg/24 h</td>
<td>17.04 ± 5.71</td>
<td>19.31 ± 1.31</td>
</tr>
<tr>
<td>Creatinine clearance, ml·min⁻¹·g⁻¹</td>
<td>0.56 ± 0.04</td>
<td>0.56 ± 0.02</td>
</tr>
<tr>
<td>kidney wt⁻¹</td>
<td>0.56 ± 0.04</td>
<td>0.56 ± 0.02</td>
</tr>
<tr>
<td>Urine osmolality, mosmol/kgH2O</td>
<td>677 ± 101</td>
<td>622 ± 47</td>
</tr>
<tr>
<td>Basal body weight, g</td>
<td>258 ± 4</td>
<td>262 ± 4</td>
</tr>
<tr>
<td>Final body weight, g</td>
<td>270 ± 6</td>
<td>272 ± 5</td>
</tr>
<tr>
<td>Food consumed, g/day</td>
<td>19.4 ± 0.9</td>
<td>19.8 ± 1.0</td>
</tr>
<tr>
<td>H2O consumed, ml/day</td>
<td>69 ± 4</td>
<td>61 ± 4</td>
</tr>
<tr>
<td>Plasma epinephrine, pg/ml</td>
<td>1,695.0 ± 182.9</td>
<td>985.5 ± 139.5*</td>
</tr>
</tbody>
</table>

Values are mean ± SE. *P < 0.05 vs. calcitriol. **P < 0.01 vs. calcitriol. ***P < 0.001 vs. calcitriol.
DISCUSSION

In this study, we have demonstrated that both calcitriol and paricalcitol increase PRA and were associated with hypercalcemia and polyuria. The paricalcitol-mediated increase in PRA was associated with elevated CaSR mRNA. We demonstrated that the calcitriol-mediated increase in PRA is eliminated by volume replacement with subcutaneous 0.9% NaCl. Additionally, the calcitriol-mediated increase in PRA was ameliorated by β-adrenergic blockade, but not COX-2 inhibition. All of these data support the thesis that vitamin D increases PRA independently of plasma Ca2+ through CaSR-mediated effects on hypercalcemia, polyuria, and volume depletion, which lead to elevated β-adrenergic activity.

It has been previously demonstrated that vitamin D analogs may increase PRA (32, 42). However, the mechanism by which this occurred was not examined, but it was always assumed that vitamin D increases PRA due to its hypercalcemic effects (32). We found that both calcitriol and paricalcitol significantly increase PRA. Importantly, we found that paricalcitol increased PRA in the absence of any changes in ionized plasma Ca2+, demonstrating that the stimulatory effects of vitamin D are not necessarily due to hypercalcemia, a significantly novel finding. It is unlikely the calcitriol- and paricalcitol-mediated increase in PRA was mediated by their inhibitory effects on PTH. PTH actually increases PRA due to its hypercalcemic effects. Plasma phosphorus is known to decrease CaSR expression (7). While we could not measure plasma phosphorus in our experiments (due to hemolysis that occurred with blood collection), we were able to measure urinary PO4− excretion, which was significantly higher in calcitriol-treated rats compared to control rats.

Values are means ± SE. **P < 0.01 vs. control, †††P < 0.001 vs. control, ‡‡‡P < 0.005 vs. control + propranolol. ††P < 0.01 vs. control + propranolol. †P < 0.05 vs. calcitriol. †††P < 0.001 vs. calcitriol.

Table 3. Effects of β-blockade with propranolol on plasma, urinary, and body weight parameters from control and calcitriol-treated rats

<table>
<thead>
<tr>
<th></th>
<th>Control</th>
<th>Control+ Propranolol</th>
<th>Calcitriol</th>
<th>Calcitriol+ Propranolol</th>
</tr>
</thead>
<tbody>
<tr>
<td>Plasma Na+, mmol/l</td>
<td>139 ± 0.4</td>
<td>139 ± 0.4</td>
<td>138 ± 0.4</td>
<td>139 ± 0.6</td>
</tr>
<tr>
<td>Plasma ionized Ca++, mmol/l</td>
<td>1.19 ± 0.02</td>
<td>1.19 ± 0.02</td>
<td>1.36 ± 0.03***</td>
<td>1.39 ± 0.04†††</td>
</tr>
<tr>
<td>Plasma osmolality, mosmol/kgH2O</td>
<td>296 ± 1</td>
<td>296 ± 2</td>
<td>308 ± 1***</td>
<td>308 ± 2†††</td>
</tr>
<tr>
<td>24-h Urine volume, ml/24 h</td>
<td>10.9 ± 1.1</td>
<td>10.1 ± 1.2</td>
<td>32.6 ± 2.8***</td>
<td>34.9 ± 3.0‡‡‡</td>
</tr>
<tr>
<td>Urinary Na+ excretion, mmol/24 h</td>
<td>1.35 ± 0.17</td>
<td>1.54 ± 0.14</td>
<td>2.28 ± 0.10***</td>
<td>1.77 ± 0.18†</td>
</tr>
<tr>
<td>Urinary Ca++ excretion, mg/24 h</td>
<td>1.20 ± 0.40</td>
<td>0.91 ± 0.21</td>
<td>10.57 ± 2.93**</td>
<td>8.15 ± 1.83‡</td>
</tr>
<tr>
<td>Creatinine clearance, ml·min−1·g kidney wt−1</td>
<td>0.48 ± 0.03</td>
<td>0.57 ± 0.05</td>
<td>0.59 ± 0.05</td>
<td>0.62 ± 0.05</td>
</tr>
<tr>
<td>Urine osmolality, mosmol/kgH2O</td>
<td>1.439 ± 157</td>
<td>1.580 ± 196</td>
<td>809 ± 46**</td>
<td>691 ± 57‡‡‡</td>
</tr>
<tr>
<td>Basal body weight, g</td>
<td>244 ± 6</td>
<td>244 ± 7</td>
<td>244 ± 4</td>
<td>243 ± 4‡‡‡</td>
</tr>
<tr>
<td>Final body weight, g</td>
<td>272 ± 6</td>
<td>270 ± 6</td>
<td>259 ± 6</td>
<td>253 ± 5‡†</td>
</tr>
<tr>
<td>Food consumed, g/day</td>
<td>17.0 ± 0.6</td>
<td>16.1 ± 0.5</td>
<td>16.2 ± 0.4</td>
<td>13.4 ± 0.6±±±</td>
</tr>
<tr>
<td>H2O consumed, ml/day</td>
<td>39 ± 3</td>
<td>39 ± 2</td>
<td>62 ± 4***</td>
<td>59 ± 2†††</td>
</tr>
</tbody>
</table>

Fig. 6. Effects of cyclooxygenase (COX)-2 inhibition with NS-398 on calcitriol-stimulated PRA. COX-2 inhibition had no effect on calcitriol-stimulated PRA. Values are means ± SE. **P < 0.01 vs. control, ‡P < 0.05 vs. control + NS-398.
calcitriol-treated rats compared with paricalcitol and controls. This suggests that the impairment of CaSR expression in calcitriol-treated rats may be due to its hyperphosphatemic effects.

Thus our data crucially demonstrate that stimulation at the VDR with paricalcitol can increase PRA independently of changes in plasma Ca\textsuperscript{2+} via hypercalcuria, polyuria, and hypovolemia. These chronic changes are in contrast to the acute effects of elevated plasma (in vivo) or media (in vitro) Ca\textsuperscript{2+} which inhibit renin release from JG cells via impaired synthesis and increased degradation of the stimulatory second messenger, cAMP (3, 4, 30, 31). Thus the effects of Ca\textsuperscript{2+}-mediated processes on renin appear to be a balance of an acute, direct inhibitory effect directly on the JG cell and a chronic, stimulatory effect via volume status.

Factors that regulate renin secretion typically do so via three different classic pathways: 1) by changes in NaCl delivery and reabsorption at the macula densa (14, 17–19); 2) by changes in renal sympathetic nerve activity acting on β-adrenergic receptors (20, 28) on the JG cells; or 3) by changes in perfusion pressure to the kidney (41, 45) via the renal baroreceptor. It has been established that the VDR is not expressed in JG cells, suggesting hypovolemia was the mechanism by which they both increased PRA. Calcitriol and paricalcitol treatment did not alter blood pressure. As such, we were unable to ascertain the role of the baroreceptor in this response. To determine whether volume depletion from the calcitriol-mediated polyuria was causing the elevated PRA, we administered sterile 0.9% NaCl subcutaneously to replace circulating volume lost from the polyuria. Volume replacement in calcitriol-treated rats completely normalized PRA, suggesting that polyuria-mediated hypovolemia is the cause by which calcitriol elevated PRA. Consistent with this result, amelioration of polyuric symptoms also normalized plasma renin levels in a patient with type-V Bartter syndrome (46). The administration of saline to the calcitriol+0.9% NaCl group returned plasma epinephrine levels to control levels without correcting them. This demonstrates that the rats treated with 0.9% NaCl were euolemic, and not volume expanded.

The elevated plasma epinephrine levels in calcitriol-treated rats suggested that elevated sympathetic activity may account for the effects of polyuria and hypovolemia on calcitriol-elevated PRA. β-Adrenergic receptor activation is a significant element in the elevation of PRA in response to water restriction and hypovolemia (20, 28). Thus it seemed plausible that the β-adrenergic pathway could mediate the response of PRA to calcitriol-mediated polyuria and hypovolemia. Moreover, elevated adrenergic activity has been reported in states of hypercalcemia as well: Patients with hyperparathyroidism have higher levels of circulating epinephrine (48). Consistent with these results, we demonstrated that β-blockade with propranolol reversed calcitriol-stimulated PRA back to control levels. Thus the sum of our data demonstrates that calcitriol elevates PRA due to polyuria-mediated hypovolemia and the resulting elevated β-adrenergic activity.

To further examine how calcitriol-mediated polyuria and hypovolemia elevates PRA, we tested whether it was due to elevated COX-2 activity, and by extension, natriuresis. Macula densa-derived COX-2 elevates PRA in response to loop diuretics or low dietary NaCl (14, 17–19). Additionally, COX-derived metabolites play an integral role in the response of the kidney to hypercalcemia: nonselective COX inhibition ameliorates vitamin D-induced polyuria (24, 38). Emerging research suggested that COX-2 may be responsible, in part, for some of these responses (49). However, our data demonstrated that COX-2 inhibition had no effect on calcitriol-stimulated PRA or polyuria. These data rule out changes in macula densa NaCl reabsorption and COX-2 activity as the cause of the elevation in PRA by calcitriol. Thus the differences in Na\textsuperscript{+} excretion seen in some of our protocols should not have been the causative factor behind the vitamin D-mediated elevations in PRA. The inability of NS-398 (a COX-2 inhibitor) to decrease PRA is not an issue of inefficient dosing, as this established dose is effective for inhibiting COX-2 (26), and lesser doses of NS-398 can also inhibit renin secretion elevated by NaCl restriction (17, 18). Moreover, our data demonstrate that the administration of NS-398 was successful, as it decreased urine osmolality in non-calcitriol-treated rats, consistent with previous results (29).

Previous data have suggested that calcitriol or other vitamin D metabolites may inhibit renin-angiotensin activity due to a direct suppression of renin transcription in the JG cell (25, 34).

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### Table 4. Effects of COX-2 inhibition with NS-398 on plasma, urinary, and body weight parameters from control and calcitriol-treated rats

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Control</th>
<th>Control + NS-398</th>
<th>Calcitriol</th>
<th>Calcitriol + NS-398</th>
</tr>
</thead>
<tbody>
<tr>
<td>Plasma Na\textsuperscript{+}, mmol/l</td>
<td>138 ± 1.3</td>
<td>138 ± 0.6</td>
<td>139 ± 0.5</td>
<td>139 ± 0.5</td>
</tr>
<tr>
<td>Plasma ionized Ca\textsuperscript{2+}, mmol/l</td>
<td>1.21 ± 0.03</td>
<td>1.21 ± 0.03</td>
<td>1.43 ± 0.03***</td>
<td>1.43 ± 0.03***</td>
</tr>
<tr>
<td>Plasma osmolality, mosmol/kgH\textsubscript{2}O</td>
<td>303 ± 2</td>
<td>302 ± 1</td>
<td>303 ± 1</td>
<td>308 ± 12†††</td>
</tr>
<tr>
<td>24-h Urine volume, ml/24 h</td>
<td>9.0 ± 1.7</td>
<td>13.0 ± 1.0</td>
<td>41.2 ± 2.6***</td>
<td>35.2 ± 3.3†††</td>
</tr>
<tr>
<td>Urinary Na\textsuperscript{+} excretion, mmol/24 h</td>
<td>1.46 ± 0.09</td>
<td>1.79 ± 0.11</td>
<td>1.79 ± 0.09</td>
<td>1.72 ± 0.08</td>
</tr>
<tr>
<td>Urinary Ca\textsuperscript{2+} excretion, mg/24 h</td>
<td>1.71 ± 0.71</td>
<td>2.44 ± 1.11</td>
<td>14.64 ± 2.06***</td>
<td>14.23 ± 2.24†††</td>
</tr>
<tr>
<td>Creatinine clearance, ml·min\textsuperscript{-1}·g kidney wt\textsuperscript{-1}</td>
<td>0.56 ± 0.06</td>
<td>0.64 ± 0.07</td>
<td>0.70 ± 0.06</td>
<td>0.73 ± 0.06</td>
</tr>
<tr>
<td>Urine osmolality, mosmol/kgH\textsubscript{2}O</td>
<td>2,103 ± 160</td>
<td>1,599 ± 71**</td>
<td>608 ± 46***</td>
<td>780 ± 78†††</td>
</tr>
<tr>
<td>Basal body weight, g</td>
<td>259 ± 3</td>
<td>261 ± 5</td>
<td>246 ± 5</td>
<td>249 ± 4</td>
</tr>
<tr>
<td>Final body weight, g</td>
<td>286 ± 6</td>
<td>287 ± 6</td>
<td>259 ± 5**</td>
<td>264 ± 4††</td>
</tr>
<tr>
<td>Food consumed, g/day</td>
<td>20.4 ± 0.8</td>
<td>20.9 ± 0.7</td>
<td>16.8 ± 0.7**</td>
<td>17.8 ± 0.7</td>
</tr>
<tr>
<td>H\textsubscript{2}O consumed, ml/day</td>
<td>43 ± 2</td>
<td>44 ± 3</td>
<td>65 ± 3***</td>
<td>61 ± 3†††</td>
</tr>
</tbody>
</table>

Values are means ± SE. COX, cyclooxygenase. **P < 0.01 vs. control. ***P < 0.001 vs. control. ‡P < 0.05 vs. control+NS-398. ‡‡P < 0.01 vs. control+NS-398. ††P < 0.01 vs. control. †††P < 0.001 vs. control+NS-398.
This concept is coming under increased scrutiny, as the VDR is not expressed in JG cells (21, 23, 51) or renin-containing As 4.1 cells (25). Thus it is unclear how vitamin D can exert a direct effect on the JG cell in the absence of a nuclear receptor. Calcitriol has been shown to decrease renal renin expression in mice (16, 25, 34). The doses (per gram body weight) used in these studies and ours are similar, and similar increases in plasma Ca\(^{2+}\) were achieved in each. Attention has been paid to mice with genetic deletions of the VDR (25) or 1-α hydroxylase (56) that develop elevated renal renin expression. This phenomenon may solely be due to the lack of cardiac VDR expression, as cardiac-specific VDR knockout mice also develop elevated renal renin expression (10), arguing against a renal-specific effect of VDR agonists on renin suppression. Our findings in a rat model showing that vitamin D can increase renin are consistent with previous data from rats and dogs (32, 42), and since calcitriol and paricalcitol increased PRA in all of our protocols, we are confident in our data.

In conclusion, we have demonstrated that calcitriol and paricalcitol both increase PRA and induce hypercalciuria and polyuria and therefore volume depletion. The increase in PRA is not due to hypercalcemia, but rather mediated by polyuria, hypervolemia, and subsequent elevated β-sympathetic activity. Our data suggest that the therapeutic strategy of treating hyperreninemic states with vitamin D analogs may be ineffective.

**Perspectives**

A potential criticism of our work is that we have used doses of calcitriol and paricalcitol that solely reflect the effects of vitamin D at levels considered toxic. However, our doses are consistent with previously validated doses in the literature (15, 16, 25, 34, 43). Conversely, we feel that our data may have clinical relevance. It has been shown that clinically relevant doses of paricalcitol induce hypercalciuria, even after only 7 days of administration in human patients (1). Moreover, three separate clinical trials have shown that paricalcitol increases plasma creatinine (1, 12, 44), which would be completely consistent with the hypovolemic effect we see in our study. At clinical doses in predialysis patients, paricalcitol decreases brain-natriuretic peptide (44), further suggesting a hypovolemic effect. Our data suggest that vitamin D analogs may affect cardiovascular parameters via changes in volume status rather than a direct effect on the renin-angiotensin system. Our data do not address whether correcting vitamin D deficiencies alters cardiovascular health, but our data suggest that treating elevated levels of renin with vitamin D may be ineffective.

**GRANTS**

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**DISCLOSURES**

No conflicts of interest, financial or otherwise, are declared by the authors.

**AUTHOR CONTRIBUTIONS**


**REFERENCES**

expression in mice deficient in beta-1 and beta-2-adrenergic receptors. 


